

UDC 577.13:582.623.2

DOI: 10.15372/CSD2019166

Phenol Compounds in the Species of *Salix* L. Genus in the World Flora

A. A. PETRUK

Central Siberian Botanical Garden, Siberian Branch of the Russian Academy of Sciences, Novosibirsk, Russia

E-mail: pet.a@mail.ru

(Received May 17, 2019; revised July 18, 2019)

Abstract

Review and analysis of the modern data on the content of phenol compounds in the plants of the *Salix* L. genus were carried out. The works aimed at the search for chemotaxonomic markers and their application in systematics are presented. It is demonstrated that the qualitative composition of phenol compounds is an invariable, stable sign for the taxons of the *Salix* genus. Phenol glycosides may be used to identify morphologically similar species and hybrid forms. Special attention is paid to the species that have been the subjects of investigations in different countries: *S. babylonica* L., *S. alba* L., *S. raddeana* Laksch. ex Nasarow, *S. caprea* L. and *S. acutifolia* Willd.

Keywords: *Salix* L., phenol compounds, flavonoids, phenol glycosides, chemotaxonomy

INTRODUCTION

The *Salix* L. genus is one of the numerous genera of Salicaceae Mirb. family, it includes 552 species of the world flora [1]. The number of species indicated for Russia and adjacent territories is 158 [2], for the Asian part of Russia – 103 species [3].

There are data on the antipyretic, disinfecting, astringent activity of osier bark both in folk and in traditional medicine [4, 5]. Research into the hemostatic, diuretic, antimicrobial, antiinflammatory and antitumour activity of the extractive substances of the plants of *Salix* genus are carried out [6–10]. Osier bark is one of the components of antirheumatic and antifebrile medicinal preparations in many countries [4]. The curative properties of osier are due to the presence of phenol compounds in the raw material [11, 12]. The information on the chemical composition of the species of *Salix* genus and their pharmacological activity was published by Zuzuk with co-

authors [4], Bontsevich [13], Khiteva [14], Frolova with co-authors [15].

The *Salix* genus is represented by three subgenera: *Salix*, *Vetrix* Dumort. and *Chamaetia* (Dumort.) Nasarow, according to the traditional classification based on the morphological and ecological-geographical characteristics of the representatives of the genus [16, 17]. The systematic position of the species, ecological-morphological characterization are presented according to the literature and electronic sources [1, 3, 16, 17]. Cycles of studies in different countries have been dedicated to separate species during the two recent decades: *S. babylonica* L., *S. alba* L., *S. raddeana* Laksch. ex Nasarow, *S. caprea* L. and *S. acutifolia* Willd. The data on the pharmacological activity were described in detail in the indicated works.

The goal of the present work was to make a review of research and analysis of modern literature data on the composition of phenol compounds in the plants of *Salix* genus, and evalua-

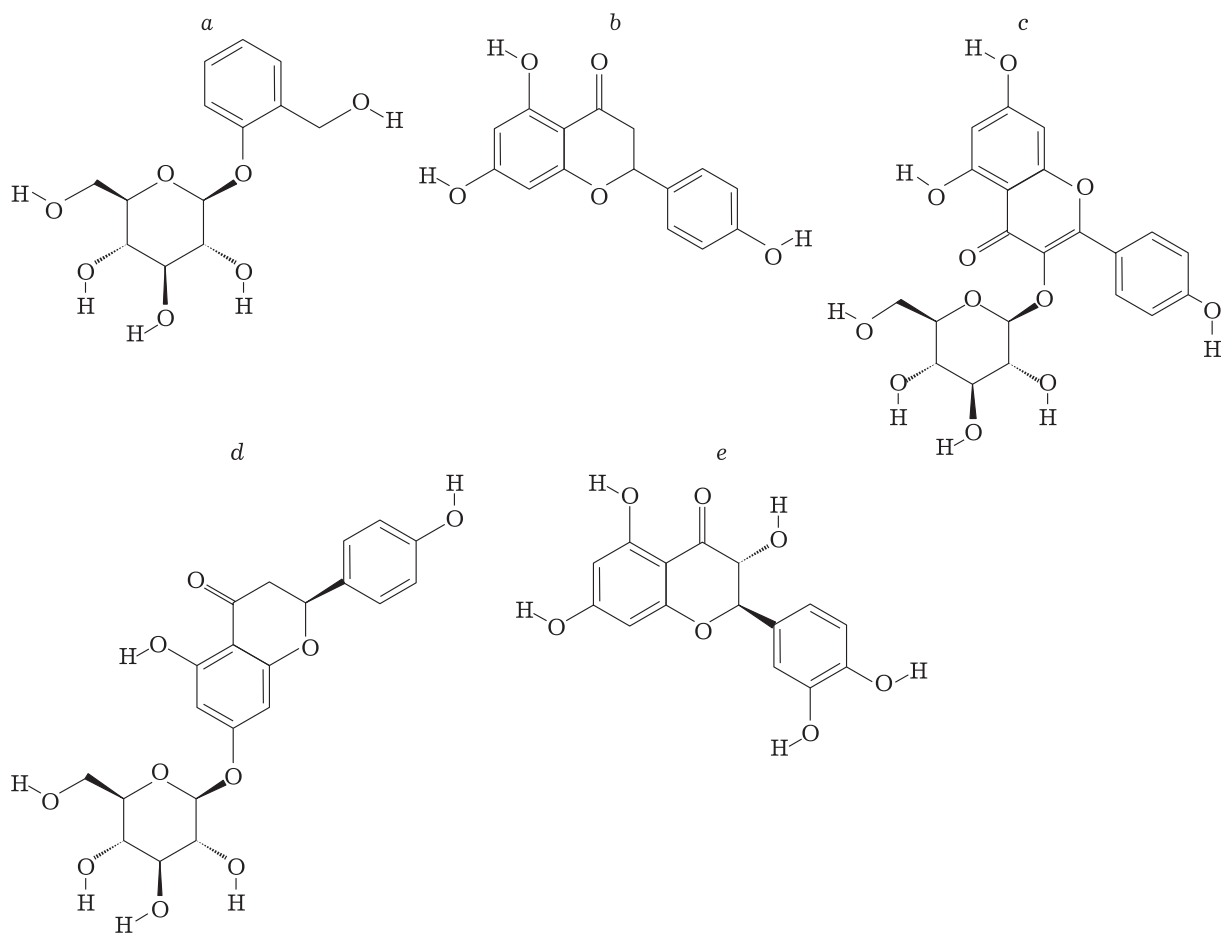


Fig. 1. The structural formulas of the main phenol compounds of *Salix* L. genus: a – salycine, b – naringenin, c – astragaline, d – prunine, e – taxifolin.

tion of the possibility to use them as chemotaxonomic markers.

PHENOL COMPOUNDS

The structural formulas of the main phenol compounds present in the plants of the *Salix* L. genus are presented in Fig. 1 [18].

Salix babylonica

S. babylonica L. (= *S. matsudana* Koidz.) is weeping willow belonging to the Subalbae Koidz. subsection of *Salix* subgenus. It is a tree up to 15 m high. The species is widespread in culture almost in all regions with moderate warmth over the globe. The origin is in arid and semi-arid regions of Central and Northern China.

Within several recent years, works dealing with the studies of composition and dynamics of phenol compounds in *S. babylonica* plants were published in China [19–25]. It was established

that total flavonoid content is higher in leaves than in branches and in blossom clusters. It reached its maximum (6.044 %) in May, and minimum (1.875 %) in July [19]. The compounds isolated from the methanol extract of the leaves of *S. babylonica* include 7-O- β -D-glucuronid of chrysoeriol, 7-O- β -D-galactopyranoside of luteolin, 7-O-glucoside of kaempferol, 7-O-galactoside of apigenin, 4'-O-glucoside of luteolin, salycine, trichocarpine, and a new phenol glycoside 2-O-acetyltrichocarpine (benzyl ester of 2-O- β -D-glucoside of gentisic acid) [20, 21]. In addition, the compounds discovered in the leaves of *S. babylonica* for the first time include β -D-glucopyranuronide of apigenine, 7-O- β -O-glucopyranuronide of luteolin and 7-O- β -D-glucopyranuronide of chrysoeriol [22]. Later a new flavonoid named matsudon was isolated, along with five known flavonoids, such as luteolin, isoquercetin, 7-methoxyflavon, 7-O-glucoside of luteolin, 4',7-dihydroxyflavon, and two phenol glycosides – leonuriside and piceoside. Their

structures were determined on the basis of extensive studies by means of 1D- and 2D-NMR, mass spectrometric analysis and comparison with literature data [23]. The content of phenol compounds in the abscised leaves of the plants of this species was studied [24]. It was shown that an increase in the time of ultrasonic treatment during the extraction of flavonoids with a 60 % aqueous solution of ethanol promotes an increase in the yield of flavonoids from 3.83 to 11.23 %. It should be stressed that the content of phenol compounds in the abscised leaves of the species of *Salix* genus growing in India was studied previously. It was discovered that the maximal amount of flavonols is characteristic of dead leaves [25].

Salix alba

S. alba L. is white willow belonging to the *Salix* section of *Salix* subgenus. It is a large tree up to 30 m high. The species is widespread in culture. The natural habitat occupies the major part of Europe; there are data on its occurrence in northern Africa, West and Middle Siberia, Middle Asia, Kazakhstan.

White willow is one of the best studied species, it is used in scientific and practical medicine [4]. The extract of the bark is known to contain eryodistiol, 5,7-dihydroxychromen-4-one and naringenine. These data were obtained using high-rate chromatography, mass spectrometry and NMR spectroscopy [26]. Investigation of water extracts from the bark of white willow and other species is carried out at the Pyatigorsk State Pharmaceutical Academy. It was revealed that the content of extractives is the highest in the extract prepared by the authors according to the XI edition of the State Pharmacopeia published in 1987 [27]. This extract of the bark of white willow possessed high antiinflammatory activity.

Salix raddeana

S. raddeana Laksch. ex Nasarow is Radde willow belonging to section *Vetrix* Dumort. of the *Vetrix* subgenus. It is a high bush or a tree up to 6–8 m high. The species is widespread in forests, forest-steppe and lower mountainous belts in Eastern Siberia, in the Far East, in the northern and north-eastern regions of Mongolia, China and Korea.

In China, several works dealt with this species [28–33]. For example, in 2004 high-performance liquid chromatography (HPLC) was applied to isolate raddeanaline (7-O-glucopyranoside of diosmetin) from *S. raddeana* and identify this compound; a procedure for more economical determination of flavons was proposed [28, 29].

Two new components for the genus were identified with the help of UV spectrophotometry and mass spectrometry: 7-O- β -L-arabinopyranosyl(1-6) β -D-glucopyranoside of luteolin and 7-O- β -D-xylopyranosyl(1-6) β -D-glucopyranoside of luteolin. For *S. raddeana*, also the following compounds were reported: 7-O- β -D-xylopyranosyl(1-6) β -D-glucopyranoside of diosmetine, rutin, astragaline and isoquercetin [30–33].

Salix caprea

S. caprea L. (= *S. hultenii* Flod.) is goat willow belonging to the *Vetrix* section of *Vetrix* subgenus. It is a tree up to 12–15 m high. It grows in forests, on clearings, at forest edges, near roads. The species is widespread over the territory of Eurasia.

The following phenol compounds were found in the wood and tree warts of goat willow: vanillic acid, catechine, galloocatechine, dihydrokaempferol, dihydromyricetin, naringenin and taxifolin. It was established that warts and branches contain higher concentrations of the substances than the stem of the same tree [34]. A procedure of an easier and rapid extraction of phenol compounds was developed for the blossom clusters of goat willow as example: an optimal version is the use of 50 % methanol, the ratio of the mass of raw material to the solvent mass should be 1 : 40, boiling time 3 h with a reflux condenser. The yield of flavonoids reached 26.82 mg/mL [35].

A comparative study of *S. caprea* and *S. repens* L. (*Vetrix* subgenus, *Incubaceae* Kerner section) species and their hybrids was carried out in 2003 in Sweden [36]. It was established that a high level of condensed tannines and low level of phenol glycosides is characteristic of *S. caprea*; for *S. repens*, quite contrary, the level of tannines is low and the level of phenol glycosides is high. Hybrids occupy an intermediate position in the content of phenol compounds. A similar investigation was also carried out for the species from North America, subgenus *Vetrix*: *S. sericea* Marshall (*Griseae* (Borrer) Barratt ex Hooker section), *S. eriocephala* Michx. (*Cordatae* Barratt ex Hooker section) and their F1 hybrids [37, 38].

The content of salicortin and 2'-cynnamoylsalicortin was determined in the leaves of *S. sericea*, and the low content of tannines in the leaves was detected. In *S. eriocephala*, these phenol glycosides were not found, but a high content of condensed tanning agents was detected in leaves. These features are inherited by hybrids, for example, F1 hybrids exhibit an intermediate value for the content of condensed tanning agents. The content of phenol glycosides is often lower than in parent forms, but this parameter is unstable and may be controlled by the recessive alleles of genes. The genetic changes in the inheritance of phenol glycosides salicortin and 2'-cynnamoylsalicortin in *S. sericea* were studied. It was demonstrated that the parameters of plant growth exhibit almost no correlation with the concentrations of these compounds; in addition, the concentration of salicortin is much higher than the concentration of 2'-cynnamoylsalicortin, the degrees of inheritance also differ: 0.2 for salicortin and 0.59 for 2'-cynnamoylsalicortin. The authors of [37, 38] concluded that phenol glycosides and condensed tanning substances in *S. sericea*, *S. eriocephala* and in their F1 hybrids form one of the genetically stable features.

Salix acutifolia

S. acutifolia Willd. is acutifoliate willow belonging to the *Daphnella* Seringe ex Duby section of *Vetrix* subgenus. It is a high bush or a tree up to 6 m in height. It grows only on open non-weedy sand. The species is widespread in Europe.

Along with white willow, this is one of the best studied species used in medical science and practice. A number of works dealing with the studies of the chemical composition of bark and leaves in the plants of this species were published [39–42]. The highest content of the sum of flavonoids was detected in *S. acutifolia* at the beginning of fruiting; by the end of vegetation it decreased nearly by a factor of 2. In comparison with acutifoliate willow, flavonoid content in the leaves of *S. triandra* L. f. *concolor* A. K. Skvortsov and *S. triandra* f. *discolor* A. K. Skvortsov is maximal at the beginning of vegetation. Flavonoid content is higher in the young leaves of male individuals is higher than in female ones. During the phase of shoot lignification, a trend for tannin accumulation was observed; a negative correlation was detected between their content in the bark of branches and the content of flavonoids in leaves [39]. In the bark of acutifoliate willow

growing in the Samara Region, 14 phenol compounds were identified by means of column chromatography, UV, IR, NMR and mass spectrometry (among these compounds, 9 were identified for the first time in this species). These compounds are: (+)-catechine, naringenine, prunine, (-)-salipurposide, isosalipurposide, 6''-cumaryl isosalipurposide, isosalipurpol, triandrin, syringin, saligenin and the products of its decomposition, acylsaligenin, salicin, salicortin and tremulacin [7, 40]. The content of salicin determined quantitatively by means of thin layer chromatography was found to vary from 3 to 10 % in the model populations of the natural growth of *S. acutifolia* in the basin of middle Volga [41]. A number of phenol metabolites were revealed by means of high-performance liquid chromatography and mass spectrometry (HPLC-MS) in *S. acutifolia* plants: salicortin, salicin, tremuloidin and a new compound – glycoside of benzoic acid named acutifoliside [42].

Other species of Salix genus

As mentioned above, investigation of the chemical composition of the extracts of bark, leaves, blossom clusters of the plants of willow genus was carried out mainly for pharmacological purposes to reveal different kinds of biological activity of the extracts and then to obtain new medicinal preparations. There are only a few works aimed at the search for chemotaxonomic markers and their application in the systematics of the *Salix* genus. For example, the flavonoid composition of 14 species of *Salix* genus growing in the arboretum of the CSBG SB RAS was studied in [43]. Various combinations of two flavons (apigenin and luteolin) and three flavons (kaempferol, quercetin and myricetin) were discovered. The plants of the same species were detected to have the same qualitative composition. The similarity of the flavonoid composition of some species was also determined, for example *S. purpurea* L. and *S. gracilistyla* Miq., *S. ledebouriana* Trautv. and *S. dasyclados* Wimm., however, it was stressed that the composition of glycosides is individual for each species.

Forster *et al.* [44] analyzed the data for the following species: *S. daphnoides* Vill., *S. pentandra* L. and *S. purpurea* growing at the territory of Poland and Germany. In the opinion of the authors, these species had clear differences in the composition of phenol glycosides. The major glycoside of *S. daphnoides* and *S. purpurea* was

salicortin, the major compound of *S. pentandra* was 2'-O-acetylsalicortin. In general, *S. daphnoides* exhibited the highest content of salicylates and phenol glycosides. The authors indicated that the content of phenol compounds in willow bark decreased during the vegetation period from March to July 2007. The investigation showed that studying the qualitative and quantitative composition of phenol glycosides it is necessary to take into account the species, a specimen and the time when the material was collected during the season [44].

A group of authors from Poland and Belgium [45–47] investigated seven species of willows growing at the territory of Poland under natural conditions and in artificial growth: *S. alba* (clone 1100), *S. daphnoides* (clone 1095), *S. acutifolia*, *S. herbacea* L., *S. viminalis* L., *S. purpurea* and *S. triandra*. The differences in the composition of phenol compounds in all species were confirmed. In addition, the presence of 29-acetylsalicortin in *S. alba* (clone 1100) was discovered for the first time.

A series of works were carried out by Riitta Julkunen-Tiitto *et al.* [47–52], as well as by a number of other international projects [53–60] dealing with the chemotaxonomic studies of phenol glycosides in the leaves, bark, branches and blossom clusters of more than 60 willow species. It was confirmed that each species possesses specific glycoside composition. The traditional classification of the genus turned out to be incompatible with the classification based only on phenol glycosides. It was pointed out that simple phenol glycosides may be used to identify morphologically similar species and hybrid forms, in combination with other indices. Investigation of the regularities of intra- and interspecific chemical variability with respect to the content of tanniferous substances and the composition of 36 phenol components was carried out with the leaves of six species (120 samples). A multidimensional cluster analysis of the data revealed that the intraspecific variations are lower on average as the interspecific ones with respect to the chemical composition. The similarity in phenol compounds weakly correlated with the phylogeny of the studied species [51].

The authors of this work studied the composition and content of phenol compounds in the species of *Salix* genus with the help of HPLC. Apigenin was found in *S. recurvigemmis* A. K. Skvortsov and *S. saxatilis* Turcz. Ex Ledeb., luteolin – in *S. acutifolia*, *S. vestita* Pursh, *S.*

berberifolia Pall., *S. myrtilloides* L., *S. saxatilis* and *S. pyrolifolia* Ledeb., *S. alba* x *S. blanda*, *S. alba* and *S. alba* var. *vitellina* (L.) Stokes. Kaempferol was identified in *S. recurvigemmis*, *S. alba* x *S. blanda* Andersson, *S. alba* and *S. alba* var. *vitellina*. Rutin was discovered only in *S. alba* and *S. alba* var. *vitellina*, quercetin – in *S. vestita*, *S. berberifolia*, *S. myrtilloides*, *S. nummularia* Andersson, *S. recurvigemmis*, *S. krylovii* E. L. Wolf, *S. sphenophylla* A. K. Skvortsov, *S. alba* x *S. blanda*, *S. alba* and *S. alba* var. *vitellina*, salicin – in *S. alba*, *S. alba* x *S. blanda* and *S. alba* var. *vitellina*. No substantial differences were detected in the composition of flavonoids between male and female plants of *S. alba*. During the vegetation period, flavonoid content in the leaves of *S. alba* var. *vitellina* varies from 0.85 to 4.60 % [61–65].

CONCLUSION

The analysis of literature data and the own studies provide evidence of the impossibility to use the quantitative content of phenol compounds as a chemotaxonomic marker for the *Salix* genus. The qualitative composition remains unchanged, genetically stable sign, so it may be used to identify the taxons. The taxons best studied on the world scale for the *Salix* genus are *S. babylonica*, *S. alba*, *S. raddeana*, *S. caprea* and *S. acutifolia*.

Acknowledgements

The work was carried out within the State Assignment for CSBG SB RAS No. AAAA-A17-117012610051-5 within the Project “Evaluation of the morphogenetic potential of the populations of plants in Northern Asia using experimental methods”.

REFERENCES

- 1 The Plant List [Electronic resource]. URL: <http://www.theplantlist.org/1.1/browse/A/Salicaceae/Salix/> (Accessed 22.01.2019)
- 2 Czerepanov S. K., Vascular Plants of Russia and Adjacent States (the former USSR), Cambridge, 1995. 516 p.
- 3 Baykov K. S. (Ed.), Synopsis of Flora of the Asian part of Russia: Vascular Plants, Novosibirsk, 2012. 640 p. (in Russ.)
- 4 Zuzuk B. M., Kutsik R. V., Nedostup A. T., Khomenets I. Z., Permyakov V. V., Fedushchak N. K., White willow *Salix alba* L. Analytical review, *Provizo*, 2005, No. 17, P. 31–36. (in Russ.)
- 5 Batorova S. M., Yakovlev G. P., Aseeva T. A., A Guide to Medicinal Plants of Traditional Tibetan Medicine,

- Novosibirsk, 2013. 292 p. (in Russ.).
- 6 Hany A., Ahmed M., Mostafa I., Sohair I., Kounosuke F., The effect of willow leaf extracts on human leukemic cells in vitro, *Journal of Biochemistry and Molecular Biology*, 2003, Vol. 36, No. 4, P. 387–389.
 - 7 Hussain H., Badawy A., Elshazly A., Elsayed A., Krohn K., Riaz M., Schulz B., Chemical constituents and antimicrobial activity of *Salix subserata*, *Records of Natural Products*, 2011, Vol. 5, No. 2, P. 133–137.
 - 8 Lysenko T. A., Khiteva O. O., Savenko I. A., Kompantseva E. V., Studies of antiinflammatory activity of aqueous extracts from bark and one-year shoots of white willow, *Vestn. Nov. Med. Tekhnologiy*, 2011, Vol. XVIII, No. 2, P. 288–290. (in Russ.).
 - 9 El-Shazly A., El-Sayed A., Fikrey E., Bioactive secondary metabolites from *Salix tetrasperma* Roxb., *Zeitschrift fur Naturforschung Biochemie, Biophysik, Biologie, Virologie*. Teil C, 2012, Vol. 67, No. 7–8, P. 353–359.
 - 10 Kompantseva E. V., Frolova O. O., Demytyeva T. M., Possibility to use weeping willow in pharmacy, *Farmatsiya i Farmakologiya*, 2013, No. 1, P. 4–7. (in Russ.).
 - 11 Budantsev A. L. (Ed.), Plant Resources of Russia: Wild Flowering Plants, Their Component Composition and Biological Activity, St. Petersburg, 2009, Vol. 2. 513 p. (in Russ.).
 - 12 Sannikova E. G., Popova O. I., Kompantseva E. V., *Salix triandra* L. – outlooks and possibilities to use it in medicine and pharmacy, *Pharmacy and Pharmacology*, 2018, Vol. 6, No. 4, P. 318–339. (in Russ.).
 - 13 Bontsevich A. I., Phytochemical investigation of acutifoliolate willow (Abstract of Candidate's Dissertation in Pharmaceutics), Samara, 2007. 22 p. (in Russ.).
 - 14 Khiteva O. O. Studies of some willow species growing at the Northern Caucasia (Abstract of Candidate's Dissertation in Pharmaceutics), Pyatigorsk, 2012. 24 p. (in Russ.).
 - 15 Frolova O. O., Kompantseva E. V., Demytyeva T. M., Biologically active substances in the plants of *Salix* genus (*Salix* L.), *Farmatsiya i Farmakologiya*, 2016, Vol. 4, Issue 2(15), P. 41–59. (in Russ.).
 - 16 Skvortsov A. K., Willows of Russia and Adjacent Countries. Taxonomical and Geographical Revision, Joensuu: University of Joensuu, 1999. 307 p.
 - 17 Bolshakov N. M., *Salicaceae* family – willows, *Flora of Siberia. Novosibirsk*, 1992, Vol. 5, P. 11–59. (in Russ.).
 - 18 Structural Formulas [Electronic resource]. URL: <https://pubchem.ncbi.nlm.nih.gov> (Accessed 17.07.2019).
 - 19 Gao W., Zhang T., Chen H., Zheng Y., Liu K., Extracting total flavone from *Salix babylonica* by microwave extraction and its dynamic content study, *Zhongguo Zhongyao Zazhi*, 2007, Vol. 32, No. 3, P. 263–265.
 - 20 Liu M.-X., Zheng Y.-N., Liu X.-F., Li X.-G., Xiang L., Isolation and identification of two flavone type compounds from *Salix matsudana*, *Chemical Research in Chinese Universities*, 1998, Vol. 14, No. 2, P. 218–220.
 - 21 Liu K., Liu H., Zhou, B., Han L., Studies on chemical constituents from *Salix babylonica* L. and their stimulation on lipolysis activity, *Journal of Fudan University (Natural Science)*, 2008, Vol. 47, No. 4, P. 520–523.
 - 22 Zheng Y.-N., Zhang J., Han L.-K., Sekiya K., Kimura Y., Okuda H., Effects of compounds in leaves of *Salix matsudana* on arachidonic acid metabolism, *Yakugaku Zasshi*, 2005, Vol. 125, No. 12, P. 1005–1008.
 - 23 Wang J.-X., Zheng S.-Z., Sun L.-P., Shen X.-W., Li Y., Studies on chemical constituents of *Salix matsudana*, *Journal of the Chinese Chemical Society*, 2002, Vol. 49, No. 3, P. 437–441.
 - 24 Wang L., Li J., Gu Y., Ultrasonic-assisted extraction of flavonoids from leaves of *Salix matsudana* Koidz., *Chemistry and Industry of Forest Products*, 2007, Vol. 27, No. S1, P. 133–136.
 - 25 Sharma V., Vaid N., Age-related changes in indolic and phenolic compounds in leaves from enforced *Salix* shoot bud, *Indian Journal of Plant Physiology*, 1997, Vol. 2, No. 3, P. 207–210.
 - 26 Du Q., Gerold J., Winterhalter P., Preparation of three flavonoids from the bark of *Salix alba* by high-speed countercurrent chromatographic separation, *Journal of Liquid Chromatography & Related Technologies*, 2004, Vol. 27, No. 20, P. 3257–3264.
 - 27 Khiteva O. O., Kompantseva E. V., Lysenko T. A., Ignatenko A. I., Aqueous extracts from white willow. Routes and forms of improvement of pharmaceutical education. Search for new physiologically active substances, 4th All-Russia Conference “Pharm. Education 2010”. Part II. “Scientific Foundations of the Development of New Medicinal Plants” (Proceedings), Voronezh, 2010, P. 391–393. (in Russ.).
 - 28 Xu C., Zheng Y., Liu T., Wang Y., RP-HPLC Determination of raddeanalin in *Salix raddeana* Laksch, *Chinese Journal of Pharmaceutical Analysis*, 2004, Vol. 24, No. 6, P. 631–632.
 - 29 Xu C., Zheng Y., Liu T., Wang Y., Determining the content of flavones in *Salix raddeana* using HPLC, *Jilin Nongye Daxue Xuebao*, 2004, Vol. 26, No. 1, P. 50–52.
 - 30 Xu C., Zhang Y., Wang J., Li X., Sun G., The structure of two flavonol glycosides from the leaves of *Salix raddeana* Laksch, *Chinese Journal of Analytical Chemistry*, 2004, Vol. 32, No. 12, P. 1663–1666.
 - 31 Xu C., Zheng Y., Yang X., Li X., Sun G., Studies on the structural identification of new flavonol glycoside of *Salix raddeana* Laksch., *Chinese Journal of Analytical Chemistry*, 2005, Vol. 33, No. 9, P. 1311–1314.
 - 32 Xu C.-L., Zheng Y.-N., Han L.-K., Okuda T., Study on stimulating fat metabolism activities of alcohol extract and flavonoid constituents from leaves of *Salix raddeana*, *Chinese Pharmaceutical Journal*, 2005, Vol. 40, P. 753–756.
 - 33 Xu C.-L., Zheng, Y.-N., Yang X.-W., Li X.-G., Li X., Chen Q.-C., Raddeanalin, a new flavonoid glycoside from the leaves of *Salix raddeana* Laksch, *Journal of Asian Natural Products Research*, 2007, Vol. 9, No 5, P. 415–419.
 - 34 Pohjamo S. P., Hemming J. E., Willfor S. M., Reunanen M. H. T., Holmbom B. R., Phenolic extractives in *Salix caprea* wood and knots, *Phytochemistry*, 2003, Vol. 63, No. 2, P. 165–169.
 - 35 Sun H., Ailaiti, S., Maiwulan M., Guli A., Study on total flavonoid content from *Salix caprea*, *Chinese Journal of Ethnomedicine and Ethnopharmacy*, 2008, Vol. 45, No. 1, P. 84–87.
 - 36 Hallgren P., Ikonen A., Hjaelten J., Roininen H., Inheritance patterns of phenolics in F1, F2, and back-cross hybrids of willows: implications for herbivore responses to hybrid plants, *Journal of Chemical Ecology*, 2003, Vol. 29, No. 5, P. 1143–1158.
 - 37 Orians C. M., Roche B. M., Fritz R. S., The genetic basis for variation in the concentration of phenolic glycosides in *Salix sericea*: an analysis of heritability, *Biochemical Systematics and Ecology*, 1996, Vol. 24, No. 7–8, P. 719–724.
 - 38 Orians C. M., Griffiths M. E., Roche B. M., Fritz R. S., Phenolic glycosides and condensed tannins in *Salix sericea*, *S. eriocephala* and their F1 hybrids: not all hybrids are created equal, *Biochemical Systematics and Ecology*, 2000, Vol. 28, No. 7, P. 619–632.
 - 39 Nikitina V. S., Orazov O. E., Dynamics of total flavonoid content in the leaves and tannides in the bark of branches of *Salix triandra* L. and *S. acutifolia* Willd. individuals of different sexes, *Rast. Resursy*, 2001, Vol. 37, No. 3. P. 65–72. (in Russ.).

- 40 Zapesochnaya G. G., Kurkin V. A., Braslavskii V. B., Filatova N. V., Phenolic compounds of *Salix acutifolia* bark, *Chemistry of Natural Compounds*, 2002, Vol. 38, No. 4, P. 314–318.
- 41 Koptina A. V., Shurgin A. I., kanarskiy A. V., Ponomareva T. A., Sergeev R. V., Salicin content in the extracts of the bark of *Salix acutifolia* L., V All-Russia Conference of Chemistry and Technology of Plant Substances (Proceedings), Syktyvkar – Ufa, 2008. P. 160. (in Russ.).
- 42 Wu Y., Dobermann D., Beale M. H., Ward J. L., Acutifoliside, a novel benzoic acid glycoside from *Salix acutifolia*, *Natural Product Research*, 2016, Vol. 30, No. 15, P. 1731–1739.
- 43 Vysochina G. I., Vstovskaya T. N., On taxon specificity of the flavonoid composition in *Salix* L. genus, *Sib. Ecol. Journ.*, 1999, No. 3, P. 245–250. (in Russ.).
- 44 Forster N., Ulrichs C., Zander M., Katzel R., Mewis I., Influence of the season on the salicylate and phenolic glycoside contents in the bark of *Salix daphnoides*, *Salix pentandra* and *Salix purpurea*, *Journal of Applied Botany and Food Quality*, 2008, Vol. 82, No. 1, P. 99–102.
- 45 Poblocka-Olech L., Nederkassel A.-M., Heyden Y. V., Krauze-Baranowska M., Glod D., Baczek T., Chromatographic analysis of salicylic compounds in different species of the genus *Salix*, *Journal of Separation Science*, 2007, Vol. 30, No. 17, P. 2958–2966.
- 46 Krauze-Baranowska M., Poblocka-Olech L., Glod D., Wiwart M., Zielinski J., Migas P., HPLC of flavanones and chalcones in different species and clones of *Salix*, *Acta Pol. Pharm.*, 2013, Vol. 70, No. 1, P. 27–34.
- 47 Sulima P., Krauze-Baranowska M., Przyborowski J. A., Variations in the chemical composition and content of salicylic glycosides in the bark of *Salix purpurea* from natural locations and their significance for breeding, *Fitoterapia*, 2017, Vol. 118, P. 118–125.
- 48 Meier B., Julkunen-Tiitto R., Tahvanainen J., Sticher O., Comparative high-performance liquid and gas-liquid chromatographic determination of phenolic glucosides in *Salicaceae* species, *Journal of Chromatography*, 1988, Vol. 442, P. 175–186.
- 49 Julkunen-Tiitto R., Phenolic constituents of *Salix*: A chemotaxonomic survey of further Finnish species, *Phytochemistry*, 1989. Vol. 28, No. 8. P. 2115–2125.
- 50 Julkunen-Tiitto R., Meier B., The enzymatic decomposition of salicin and its derivatives obtained from *Salicaceae* species, *Journal of Natural Products*. 1992. Vol. 55, No. 9. P. 1204–1212.
- 51 Nyman T., Julkunen-Tiitto R., Chemical variation within and among six northern willow species, *Phytochemistry*, 2005, Vol. 66, No. 24, P. 2836–2843.
- 52 Lavola A., Maukonen M., Julkunen-Tiitto R., Variability in the composition of phenolic compounds in winter-dormant *Salix pyrolifolia* in relation to plant part and age, *Phytochemistry*, 2018, Vol. 153, P. 102–110.
- 53 Julkunen-Tiitto R., Virjamo V., Biosynthesis and roles of Salicaceae salicylates, in: Plants specialized metabolism. Genomics, biochemistry and biological functions, Arimura G., Maffei M. (Eds), Taylor & Francis Group, BocaRaton, 2017, P. 65–83.
- 54 Brereton N. J. B., Berthod N., Lafleur B., Pedneault K., Pitre F. E., Labrecque M., Extractable phenolic yield variation in five cultivars of mature short rotation coppice willow from four plantations in Quebec, *Industrial Crops and Products*, 2017, Vol. 97, P. 525–535.
- 55 Corradi E., Schmidt N., Raber N., De Mieri M., Hamburger M., Butterweck V., Potterat O., Metabolite profile and antiproliferative effects in HaCaT cells of a *Salix reticulata* extract, *Planta Med.*, 2017, Vol. 83 (14/15), P. 1149–1158.
- 56 Ruuhola T., Nybakken L., Randriamanana T., Lavola A., Julkunen-Tiitto R., Effects of long-term UV-exposure and plant sex on the leaf phenoloxidase activities and phenolic concentrations of *Salix myrsinifolia* (Salisb.), *Plant Physiology Biochemistry*, 2018, Vol. 126, No. 2, P. 55–62.
- 57 Nissinen K., Virjamo V., Mehtdtalo L., Lavola A., Valtonen A., Nybakken L., Julkunen-Tiitto R., A Seven-year study of phenolic concentrations of the dioecious *Salix myrsinifolia*, *Journal of Chemical Ecology*, 2018, Vol. 44, No. 4, P. 416–430.
- 58 Noleto-Dias C., Ward J. L., Bellisai A., Lomax C., Beale M. H., Salicin-7-sulfate: A new salicinoid from willow and implications for herbal medicine, *Fitoterapia*, 2018, Vol. 127, P. 166–172.
- 59 Noleto-Dias C., Wu Y., Bellisai A., Macalpine W., Beale M. H., Ward J. L., Phenylalkanoic glycosides (non-salicinoids) from wood chips of *Salix triandra* x *dasyclados* hybrid willow, *Molecules*, 2019, Vol. 24 (6), P. 1–12.
- 60 Tyskiewicz K., Konkol M., Kowalski R., Roj E., Warminski K., Krzyzaniak M., Gil L., Stolarski M. J., Characterization of bioactive compounds in the biomass of black locust, poplar and willow, *Trees – Structure and Function*, 2019, Vol. 33, No. 2, P. 1–29.
- 61 Petruk A. A., Phenol compounds in some representatives of *Salix* genus (Salicaceae) in the Asian part of Russia, *Khimiya Rast. Syrya*, 2011, No. 4, P. 181–185. (in Russ.).
- 62 Petruk A. A., Seasonal dynamics of changes in flavonoid and tanning substances in leaves of blossom clusters of *Salix alba* (Salicaceae), *Rast. Mir Aziat. Rossii*, 2012, No. 1, P. 72–76. (in Russ.).
- 63 Petruk A. A., Investigation of the composition of flavonoids in *Salix alba* and *S. alba* var. *vitellina* by means of HPLC, *Khimiya Rast. Syrya*, 2012, No. 2, P. 151–154. (in Russ.).
- 64 Petruk A. A., Reention of salicin in the leaves of blossom clusters of some species of *Salix* genus (Salicaceae), *Vestn. Tambov. Un-ta. Ser. Estestv. i Tekhn. Nauki*, 2013, Vol. 18, No. 3, P. 825–826. (in Russ.).
- 65 Petruk A. A., Studies of the composition and dynamics of flavonoid retention in *Salix alba* x *Salix blanda* by means of HPLC, *Voprosy Biolog., Med. i Farmatsevt. Khimii*, 2014, No. 4, P. 61. (in Russ.).